This article was downloaded by: On: 23 January 2011 Access details: Access Details: Free Access Publisher Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713617200

Preparation of 1-O-Acyl-D-Glycopyranoses via Chloroacetylated

Glycopyranosyl Donors Thomas Ziegler^a; Guido Pantkowski^a ^a Institute of Organic Chemistry, University of Stuttgart Pfaffenwaldring 55, Stuttgart, Germany

To cite this Article Ziegler, Thomas and Pantkowski, Guido(1993) 'Preparation of 1-*O*-Acyl-D-Glycopyranoses *via* Chloroacetylated Glycopyranosyl Donors', Journal of Carbohydrate Chemistry, 12: 3, 357 — 370 To link to this Article: DOI: 10.1080/07328309308018996 URL: http://dx.doi.org/10.1080/07328309308018996

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

PREPARATION OF 1-O-ACYL-D-GLYCOPYRANOSES

VIA CHLOROACETYLATED GLYCOPYRANOSYL DONORS

Thomas Ziegler * and Guido Pantkowski

Institute of Organic Chemistry, University of Stuttgart Pfaffenwaldring 55, D-W-7000 Stuttgart 80, Germany

Received September 1, 1992, Final Form February 3, 1993

ABSTRACT

Various 2,3,4,6-tetra-O-chloroacetyl-D-glucopyranosyl donors 2 were coupled to cinnamic acid (3a) and silver cinnamate (4a) to give the corresponding chloroacetylated 1-O-cinnamoyl-glucopyranose 5a in good yields. The highest α -selectivity ($\alpha/\beta = 87:13$) was achieved by coupling of the α -fluoride 2c with 3a under boron trifluoride catalysis whereas the α -bromide 2g afforded exclusively β -5a upon treatment with 4a. Thus, the D-glucosyl donors 2c and 2g and 2,3,4,6-tetra-O-chloroacetyl- α -D-galactopyranosyl bromide (6) were condensed with a series of acids 3a-e (cinnamic, 2-methyl-4-pentenoic, 3,4,5-trimethoxybenzoic, oleic, and 4-chloroacetoxy cinnamic acid, respectively) and silver salts 4a-d thereof. From α/β -mixtures 5a-c and 5e obtained from 2c and the respective acid 3, the α -products were isolated in moderate yield. Selected examples of thus prepared 1-O-acyl-D-glucoses 5 and D-galactoses 7 were dechloroacetylated using 1-piperidinethiocarbamide to give the corresponding unblocked 1-O-acyl monosaccharides 8 and 9 in 52-64% yield.

INTRODUCTION

Recently, we applied easily accessible fully chloroacetylated D-glycopyranosyl bromides for the convenient preparation of some base and hydrogenolysis-sensitive D-glycosides.¹ The use of chloroacetyl groups for the temporary protection of these derivatives is promising because dechloroacetylation can be done under essentially neutral

conditions,^{1,2} leaving labile aglycons intact. Especially for the preparation of the title compounds this synthetic approach via chloroacetylated glycosyl halides appeared particularly useful. Since 1-O-acyl-D-glycopyranoses are widespread naturally occurring sugars (for example tannins) that also embrace derivatives with significant physiological properties,³ several methods have been developed so far for their preparation. Most notably, 1-O-acyl-glycoses were prepared by regioselective acylation of the anomeric position of unprotected mono and disaccharides.^{4,5} Other, less generally applicable, methods used partially trifluoroacetyl⁶ and triethylsilyl⁷ protected derivatives, respectively for the introduction of an acyl substituent at the anomeric position and the following hydrolytic deblocking. A gibberellin- β -D-glucosyl ester was obtained by enzymatic saponification of its synthetic peracetylated precursor, however, in poor yield and thus, seemed not to be generally suitable for preparative purposes. Recently, 2,3,4,6-tetra-Ochloroacetyl-D-glycopyranoses have been esterified at the anomeric hydroxyl by a Mitsunobu protocol followed by dechloroacetylation to give the corresponding 1-O-acyl derivatives,⁸ however, with rather low α/β -selectivity. We have extended our previous approach via chloroacetylated glycopyranosyl halides for the preparation of various 1-Oacyl glycoses, since we expected this method to provide the title compounds with high diastereoselectivity.

RESULTS AND DISCUSSION

In order to determine the stereoselectivity of the formation of 1-O-acyl glycopyranoses, various 2,3,4,6-tetra-O-chloroacetyl-D-glycopyranosyl donors 2 and 6 were prepared as follows, and were tested in reactions with cinnamic acid 3a and the silver salt 4a thereof. The donors 2a-c were obtained by chloroacetylation of the corresponding glucosides **1a-c** using chloroacetic anhydride.^{1,9} The trichloroacetimidate **2f** was prepared via accordingly synthesized^{1,10} benzyl 2,3,4,6-tetra-O-chloroacetyl-β-D-glucopyranoside (2d), hydrogenolysis of which afforded first 2,3,4,6-tetra-O-chloroacetyl-D-glucopyranose (2e). The latter compound, also conveniently available⁸ by treatment of 1,2,3,4,6-penta-O-chloroacetyl- α -D-glucopyranose^{1,8} with hydrazine acetate, afforded the trichloroacetimidate 2f upon reaction with trichloroacetonitrile. The bromides 2g and 6 were prepared as described previously.¹ Table 1 summarizes the results of the condensation of the donors 2 with cinnamic acid 3a (for 2a-c and 2f) and its silver salt 4a (for 2g and 6). respectively. In all cases, the previous finding^{1,11} that chloroacetyl groups present in glycosyl donors decrease their reactivity towards nucleophiles was also evident here. For example, all chloroacetylated donors 2 needed several hours at room temperature in order to give a complete reaction. The phenyl 1-thio-D-glucoside (2a) did not react at all. Only

| 2 | 3,4 | conditions ^a | yield | product | α/β-ratio ^b |
|------------|------------|--|--------|---------|------------------------|
| 2a | 3a | NIS, 10 mol-% TfOH, 3 d | traces | 5a | _ |
| 2 b | 3a | NIS, 10 mol-% TfOH, 2 h | 77% | 5a | 30:70 |
| 2 c | 3a | 5 equiv. BF ₃ OEt ₂ , 32 h | 58% | 5a | 81:19° |
| 2 f | 3 a | 10 mol-%, TMSOTf, 1.5 h | 75% | 5a | 8:92 |
| 2 g | 4a | CH ₂ Cl ₂ , 30 h | 71% | 5a | 0:100 |
| 6 | 4 a | CH_2Cl_2 , 30 h | 70% | 7a | 0:100 |

Table 1. Reaction of various 2,3,4,6-tetra-O-chloroacetyl-D-glycopyranosyl donors 2 and 6 with cinnamic acid 3a and the silver salt 4a thereof at room temperature.

a. NIS: *N*-iodosuccinimide, TfOH: trifluoromethanesulfonic acid, TMSOTf: trimethylsilyl trifluoromethanesulfonate. b. Determined by ¹H NMR. c. α -5a was isolated from the mixture in 39% yield.

traces of product 5a could be detected on TLC. Aryl 1-thio-glycosides are usually less reactive under thiophilic activation than alkyl 1-thio-glycosides. However, phenyl 2,3,4,6-tetra-O-acetyl-1-thio- β -D-glucopyranoside does react well with nucleophiles when activated with NIS/TfOH.¹² Obviously, the chloroacetyl groups present in 2a deactivated this donor to such an extent that no reaction with 3a occurred, whereas 2b was reactive enough to give 5a in a good yield. Even the imidate 2f showed a low reactivity although imidates are usually rather reactive donors. Nevertheless, yields of chloroacetylated 1-O-acyl-D-glycosides 5a and 7a were good when prolonged reaction times were applied.



| $\begin{array}{c ccccccccccccccccccccccccccccccccccc$ | _ | | | | | | |
|---|---|-----|-----|--------------------------------------|-------|---------|------------------------------------|
| $\begin{array}{c ccccccccccccccccccccccccccccccccccc$ | | 2,6 | 3,4 | R ¹ | yield | product | α/β -ratio ^a |
| $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | | 2 c | 3b | MeQ | 52% | 5b | 90:10 ^b |
| $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | | 2 g | 4b | MeO- | 64% | 5b | 0:100 |
| $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | | 6 | 4b | MeO | 56% | 7 b | 0:100 |
| $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | | 2 c | 3c | | 60% | 5 c | 72:28 ^b |
| $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | | 2 g | 4 c | H ₂ C, CH ² CH | 70% | 5 c | 0:100 |
| $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | | 6 | 4 c | CH ₃ | 55% | 7 c | 0:100 |
| $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | | 2 c | 4d | | _c | 5d | _c |
| $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | | 2 g | 4d | | 75% | 5 d | 0:100 |
| | | 6 | 4d | | 69% | 7 d | 0:100 |
| | | 2 c | 3e | CIACO-CIACO-CH=CH- | 51% | 5e | 77:23 ^b |

Table 2. Reaction of 2,3,4,6-tetra-O-chloroacetyl- α -D-glycopyranosyl halides 2c, 2g, and 6 with various carboxylic acids 3 and silver salts 4 thereof.

a. Determined by ¹H NMR. b. From anomeric mixtures α -5 was isolated in the following yields: α -5b (36%), α -5c (34%), α -5e (27%). c. Decomposition of compound 4d.

The stereoselective outcome of the condensation depended strongly on the particular donor and the promotor used for its activation. The heterogenous reactions of the bromides 2g and 6 with silver cinnamate 4a in dichloromethane resulted expectedly¹ in the exclusive formation of the chloroacetylated 1-O-cinnamoyl- β -D-glycopyranoses 5a and 7a, respectively. Other solvents were not suitable because reaction of the chloroacetyl groups can occur.¹ In contrast, under homogenous conditions (*i.e.*, in reactions where the free acid 3a was applied) a significant amount of the α -anomer of 5a was formed. Since these condensations were performed under acidic and rather thermodynamic conditions due to the relatively long reaction times it is likely that the β -anomers were formed initially¹ with subsequent anomerisation to give the corresponding α -anomers. For preparative purposes, however, a high α/β -ratio was desirable because chromatographic separation of the anomers was difficult and a low β -content faciliated the purification of the α -anomers. The content of the α -anomer and of the yield of 5a could not be increased by using other solvents or other Lewis acids (no further details). Therefore, we applied the glucosyl fluoride 2c under BF₃-catalysis and the bromides 2g and 6 in dichloromethane for further preparations of chloroacetylated 1-O-acyl-D-glycopyranoses (Table 2).

| 5, 7 | β -5a | β -5b | β -5c | β -5d | β- 7a | β -7b | |
|---------|--------------|--------------|--------------|--------------|--------------|--------------|--|
| yield | 58% | 61% | 56% | 52% | 64% | 55% | |
| product | 8a | 8 b | 8 c | 8 d | 9a | 9b | |

Table 3. Deblocking of selected 1-*O*-acyl-2,3,4,6-tetra-*O*-chloroacetyl-D-glycopyranoses 5 and 7 with 1-piperidinylthiocarbamide.



For the dechloroacetylation of sugar derivatives thiourea^{1,10} and hydrazinedithiocarbonate^{1,2,8} are usually applied. However, since we previously encountered some problems when using these reagents for the deblocking of fully chloroacetylated glycosides, due to difficulties in removing excess of deblocking reagent,¹ we here tested several *N*,*N*-dialkyl thiourea derivatives for the final preparation of 1-*O*-acylglycopyranoses. It turned out that best results with respect to yield and ease of purification of the deblocked compounds were obtained with 1-piperidinethiocarbamide.¹³ The latter thiourea derivative has been successfully applied for the high-yielding removal of *N*chloroacetyl protective groups in amino acids.¹⁴ Table 3 shows some representative results for the dechloroacetylation of selected derivatives **5** and **7** to give the title compounds **8** and **9** in moderate to good yield. A particular advantage of 1-piperidinethiocarbamide was that the reagent and the pseudothiohydantoin formed from the latter and the chloroacetic esters could be conveniently removed by a single chromatography.



EXPERIMENTAL

General Methods. NMR data were extracted from spectra measured in solutions of CDCl₃ for blocked compounds and of CD₃OD for deblocked compounds (with TMS as an internal standard) at 25 °C with a Bruker AC 250F spectrometer. Proton-signal assignments were made by first order analysis of the spectra. Of the two magnetically nonequivalent geminal protons at C-6 of the monosaccharides, the one resonating at lower field was donated H6a and the one resonating at higher field was donated H6b. Carbon-signal assignments were made by mutual comparison of the spectra and by comparison with spectra of related compounds.^{1,8,10} Optical rotations were measured at 25 $^{\circ}$ C with a Perkin-Elmer automatic polarimeter, Model 241. Melting points were measured with a Büchi apparatus, Model SMP-20. Thin-layer chromatography (TLC) was performed on precoated plastic sheets, Polygram SIL UV254, 40 x 80 mm (Macherey-Nagel) using appropriately adjusted mixtures of carbon tetrachloride-acetone, dichloromethane-methanol and toluene-acetone, respectively for the developing. Detection was effected with UV light, where applicable, by iodine, and by charring with 5% sulfuric acid in ethanol. Preparative chromatography was performed by elution from columns of Silica Gel 60 (Merck) using solvent mixture A, carbon tetrachloride-acetone and B, dichloromethanemethanol. For the chromatography of the deblocked compds 8 and 9 the silica gel was successively prewashed with water, methanol, acetone and *n*-hexane and dried at 120 °C. Solutions in organic solvents were dried with anhydrous sodium sulfate, and concentrated at 2 kPa, ≤ 40 °C. Silver salts 4a-d, were prepared by adding an equimolar amount of aqueous silver nitrate solution to a solution of the corresponding acids 3a-d and an equimolar amount of sodium hydroxide in water. The precipitated silver salts were collected by filtration, washed with ice-cold water and dried in vacuo.

Phenyl 2, 3, 4, 6-Tetra-O-chloroacetyl-1-thio-β-D-glucopyranoside (2a). Sodium hydrogen carbonate (2.4 g. 28.6 mmol) was added at room temp to a stirred solution of phenyl 1-thio-β-D-glucopyranoside (6.0 g, 22.0 mmol; prepared by deacetylation¹⁵ of the corresponding acetylated derivative¹⁶) and chloroacetic anhydride (23.0 g, 0.134 mol) in DMF (40 mL). The mixture was stirred for 3 h and poured into water (250 mL). The solid that separated was collected by filtration, washed with water and dried in *vacuo*. Crystallisation from acetone/*n*-hexane gave 2a (11.4 g, 89%), mp 130 °C, [α]_D -10.0° (*c* 1.0, CHCl₃). ¹H NMR δ 5.36 (t, 1 H, $J_{2,3} = J_{3,4} = 9.3$ Hz, H-3), 5.12 (t, 1 H, $J_{4,5} = 9.7$ Hz, H-4), 5.02 (dd, 1 H, $J_{1,2} = 10.0$ Hz, H-2), 4.76 (d, 1 H, H-1), 4.42-4.26 (m, 2 H, H-6a,6b), 4.09, 4.08, 4.01, 3.97 (s, 3 x d, 4 x 2 H, CH₂Cl), 3.86 (ddd, 1 H, $J_{5,6a} = 4.3$ Hz, $J_{5,6b} = 2.8$ Hz, H-5); ¹³C NMR δ 85.1 (C-1), 75.2, 75.0 (C-5,3), 70.9 (C-2), 69.3 (C-4), 63.2 (C-6), 40.6, 40.4, 40.3, 40.2 (CH₂Cl). Anal. Calcd for C₂₀H₂₀Cl₄O₉S: C, 41.54; H, 3.49; Cl, 24.52; S, 5.55. Found: C, 41.61; H, 3.48; Cl, 24.77; S, 5.81.

Ethyl 2,3,4,6-Tetra-O-chloroacetyl-1-thio-β-D-glucopyranoside (2b). Sodium hydrogen carbonate (6.0 g, 71.5 mmol) was added at room temp to a stirred solution of ethyl 1-thio-β-D-glucopyranoside (1.7 g, 7.6 mmol; prepared by deacetylation of the corresponding acetylated derivative¹⁷) and chloroacetic anhydride (6.8 g, 40.0 mmol) in DMF (50 mL). The mixture was stirred for 2 h, poured into water (250 mL) and extracted with dichloromethane. The combined organic layers were washed with water and concentrated. Chromatography (solvent A, 5:1) of the residue gave 2b (3.47 g, 86%) as a colourless oil, $[\alpha]_D$ -16.6° (*c* 1.3, chloroform).¹H NMR δ 5.37 (t, 1 H, $J_{3,4}=J_{4,5}=9.4$ Hz, H-4), 5.81 (bt, 1 H, $J_{1,2}=10.1$ Hz, $J_{2,3}=9.6$ Hz, H-2), 5.12 (t, 1 H, H-3), 4.58 (d, 1 H, H-1), 4.36-4.34 (m, 2 H, H-6a,6b), 4.03, 4.12, 4.06, 4.00 (3 x s, d, 4 x 2 H, CH₂Cl), 3.98-3.81 (m, 1 H, H-5), 2.83-2.26 (m, 2 H, SCH₂CH₃), 1.28 (t, J = 7.5 Hz, 3 H, SCH₂CH₃); ¹³C NMR δ 83.2 (C-1), 75.3, 74.9, 70.9, 69.5 (C-2,3,4,5), 63.3 (C-6), 40.6, 40.4, 40.2 (2 x 1 C, 2 C, CH₂Cl), 24.3 (SCH₂CH₃), 14.8 (SCH₂CH₃).

Anal. Calcd for C₁₆H₂₀Cl₄O₉S: C, 36.25; H, 3.80; Cl, 26.75; S, 6.05. Found: C, 36.36; H, 3.88; Cl, 26.93; S, 5.95.

2,3,4,6-Tetra-*O*-chloroacetyl- α -D-glucopyranosyl Fluoride (2c). Sodium hydrogen carbonate (2.4 g, 28.6 mmol) was added at room temp to a stirred solution of α -D-glucopyranosyl fluoride (4.0 g, 22.0 mmol) and chloroacetic anhydride (21.0 g, 0.123 mol) in DMF (40 mL). The mixture was stirred for 1 h and poured into water (250 mL). The solid that separated was collected by filtration, washed with water and dried in *vacuo*. Crystallisation from acetone/diethyl ether gave 2c (8.25 g, 80%), mp 122 °C, [α]_D +69.6° (*c* 1.0, CHCl₃). ¹H NMR δ 5.80 (dd, 1 H, $J_{1,2} = 2.7$ Hz, $J_{1,F} = 52.6$ Hz, H-1), 5.60 (t, 1 H, $J_{2,3} = J_{3,4} = 9.8$ Hz, H-3), 5.25 (t, 1 H, $J_{4,5} = 9.8$ Hz, H-4), 5.06 (ddd, 1 H, $J_{2,F} = 23.5$ Hz, H-2), 4.43-4.26 (m, 3 H, H-5,6a,6b), 4.13, 4.10, 4.03, 4.00 (2 x s, 2 x d, 4 x 2 H, CH₂Cl); ¹³C NMR δ 103.2 (d, $J_{C,F} = 231.0$ Hz, C-1), 71.2 (d, $J_{C,F} = 24.7$ Hz, C-2), 70.6 (C-3), 69.3 (d, $J_{C,F} = 4.3$ Hz, C-5), 68.4 (C-4), 62.4 (C-6), 40.5, 40.24, 40.21, 40.17 (CH₂Cl).

Anal. Calcd for C₁₄H₁₅Cl₄FO₉: C, 34.45; H, 3.10; Cl, 29.06. Found: C, 34.61; H, 3.16; Cl, 29.02.

2,3,4,6-Tetra-O-chloroacetyl-D-glucopyranosyl Trichloroacetimidate (2f). A mixture of 2e (2.43 g, 5.0 mmol; prepared from compound 2d as described^{1,10}), trichloroacetonitrile (2.0 mL) and potassium carbonate (2.0 g) in dichloromethane (15 mL) was stirred at room temp for 4 h and filtered through a layer of Celite. Concentration and chromatography (solvent A, 5:1) of the residue gave a 2.6:1 α/β -mixture (¹H NMR) of 2f (1.89 g, 60%) as a colourless foam, $[\alpha]_D$ +54.8° (c 0.5, chloroform). ¹H NMR (significant peaks) α -2f: δ 6.60 (d, 1 H, $J_{1,2} = 3.7$ Hz, H-1), 5.68 (t, 1 H, $J_{2,3} = J_{3,4} = 9.8$ Hz, H-3), 5.26 (dd, 1 H, H-2); β -2f: δ 6.01 (d, 1 H, $J_{1,2} = 5.5$ Hz, H-1), 5.25 (dd, 1 H, $J_{2,3} = 9.6$ Hz, H-2), 4.76 (ddd, 1 H, $J_{5,6a} = 5.3$ Hz, $J_{5,6b} = 3.0$ Hz, H-5); ¹³C NMR α -2f: δ 160.5 (C=NH), 116.0 (CCl₃), 92.3 (C-1), 73.5, 71.1, 70.8 (C-2,3,5), 68.9 (C-4), 63.5 (C-6), 40.6, 40.5, 40.4, 40.3 (CH₂Cl); β -2f: δ 161.7 (C=NH), 116.4 (CCl₃), 100.5 (C-1), 77.4 (C-5), 72.1, 71.3 (C-2,3), 68.2 (C-4), 64.1 (C-6), 40.7, 40.6 (2 x 2 C, CH₂Cl).

Anal. Calcd for C₁₆H₁₆Cl₇NO₁₀: C, 30.48; H, 2.56; Cl, 39.36; N, 2.22. Found: C, 30.28; H, 2.56; Cl, 39.24; N, 2.47.

4-Chloroacetoxycinnamic Acid (3e). Chloroacetyl chloride (7.5 g, 66.4 mmol) was added at 0 °C to a stirred solution of *p*-coumaric acid (4.0 g, 24.4 mmol) in aqueous sodium hydroxide solution (3 N, 35 mL) and the resulting suspension was stirred for 5 min. After addition of aqueous HCl solution, the material that separated was collected by filtration and washed with ice-cold water. Crystallisation from acetone/*n*-hexane gave 3e (3.96 g, 68%), mp 186-187 °C. ¹H NMR (d₆-acetone) δ 7.79-7.75, 7.29-7.26 (2 x m, 2 x 2 H, H_{arom}), 7.70 (d, 1 H, J = -16.0 Hz, CH=CHCOO), 6.54 (d, 1 H, CH=CHCOO), 4.59 (s, 2 H, CH₂Cl).

Anal. Calcd for C₁₁H₉ClO₄: C, 54.90; H, 3.77; Cl, 14.73. Found: C, 55.09; H, 3.78; Cl, 14.50.

2,3,4,6-Tetra-O-chloroacetyl-1-O-cinnamoyl-D-glucopyranose (5a). (a) A mixture of **2a** (1.33 g, 2.3 mmol), **3a** (0.37 g, 2.5 mmol), *N*-iodosuccinimide (0.67 g, 3.4 mmol) and trifluoromethanesulfonic acid (60μ L, 0.7 mmol) in dichloromethane (20 mL) was stirred at room temp for 3 d. TLC (solvent *A*, 5:1) revealed the formation of traces of **5a**.

(b) A mixture of 2b (392.4 mg, 1.0 mmol), 3a (177.8 mg, 1.2 mmol), *N*-iodosuccinimide (270.0 mg, 1.2 mmol) and trifluoromethanesulfonic acid (10 μ L, 0.1 mmol) in dichloromethane (5 mL) was treated as described for (a). After stirring at room temp for 2 h the mixture was washed with aqueous sodium hydrogen carbonate and sodium thiosulfate solution. Concentration and filtration over a short column of silica gel gave a 30:70 α/β -mixture (¹H NMR) of 5a (370 mg, 77%) as a colourless foam.

(c) A solution of 2c (1.29 g, 2.63 mmol), 3a (0.48 g, 3.21 mmol) and boron trifluoride etherate (0.4 mL, 13.2 mmol) in dichloromethane (15 mL) was stirred at room temp for 32 h, washed with aqueous sodium hydrogen carbonate, concentrated and filtered through a short column of silica gel to give a 81:19 α/β -mixture (¹H NMR) of 5a (0.94 g, 58%) as a colourless foam. Chromatography (solvent A, 5:1) gave α -5a (0.64 g, 39%), [α]_D +109.3° (c 1.0, chloroform). ¹H NMR δ 7.81 (d, 1 H, J = -16.0 Hz, CH=CHCOO), 6.53 (d, 1 H, CH=CHCOO), 6.50 (d, 1 H, J_{1,2} = 3.9 Hz, H-1), 5.69 (t, 1

H, $J_{2,3} = J_{3,4} = 9.8$ Hz, H-3), 5.28 (t, 1 H, $J_{4,5} = 9.8$ Hz, H-4), 5.27 (dd, 1 H, H-2), 4.40 (dd, 1 H, $J_{5,6a} = 4.4$ Hz, $J_{6a,6b} = -12.9$ Hz, H-6a), 4.34-4.29 (m, 2 H, H-5,6b), 4.02, 4.05, 4.02, 4.01 (d, 3 x s, 4 x 2 H, CH₂Cl); ¹³C NMR δ 148.0 (*C*H=CHCOO), 115.8 (CH=CHCOO), 88.6 (C-1), 71.2, 70.4, 69.4, 69.2 (C-2,3,4,5), 62.8 (C-6), 40.5, 40.2 (2 x 2 C, CH₂Cl).

Anal. Calcd for C₂₃H₂₂Cl₄O₁₁: C, 44.83; H, 3.60; Cl, 23.01. Found: C, 45.40; H, 3.71; Cl, 22.31.

(d) Trimethylsilyl trifluoromethanesulfonate (18 μ L, 0.1 mmol) was added at -20 °C to a solution of 2f (630.5 mg, 1.0 mmol) and 3a (177.8 mg, 1.2 mmol) in dichloromethane (5 mL). The solution was stirred at room temp for 1.5 h and washed with aqueous sodium hydrogen carbonate solution. Concentration and filtration over a short column of silica gel gave a 8:92 α/β -mixture (¹H NMR) of 5a (360 mg, 75%) as a colourless foam.

(e) A suspension of 2g (887.0 mg, 1.62 mmol) and 4a (603.4 mg, 2.37 mmol) in dichloromethane (8 mL) was stirred at room temp for 30 h. The mixture was filtered through a layer of Celite and the filtrate was washed with aqueous sodium thiosulfate solution. Concentration and chromatography (solvent A, 5:1) of the residue gave β -5a (711.4 mg, 71%) as a colourless foam, $[\alpha]_D$ -4.4° (*c* 0.9, chloroform). ¹H NMR δ 7.76 (d, 1 H, J = -16.0 Hz, PhCH=CH-), 6.41 (d, 1 H, PhCH=CH-), 5.93 (d, 1 H, $J_{1,2} = 8.1$ Hz, H-1), 5.46 (t, 1 H, $J_{3,4} = J_{4,5} = 9.4$ Hz, H-4), 5.34 (dd, 1 H, $J_{2,3} = 9.3$ Hz, H-2), 5.27 (t, 1 H, H-3), 4.43 (dd, 1 H, $J_{5,6a} = 4.2$ Hz, $J_{6a,6b} = -12.6$ Hz, H-6a), 4.32 (dd, 1 H, $J_{5,6b} = 2.3$ Hz, H-6b), 4.27-4.06 (m, 1 H, H-5), 4.05, 4.13, 4.03, 4.02 (d, 3 x s, 4 x 1 H, CH₂Cl); ¹³C NMR δ 148.1 (CH=CHCOO), 115.7 (CH=CHCOO), 91.4 (C-1), 73.8, 72.1, 71.4 (C-2,3,5), 69.1 (C-4), 62.8 (C-6), 40.6, 40.2 (1 C, 3 C, CH₂Cl).

Anal. Calcd for C₂₃H₂₂Cl₄O₁₁: C, 44.83; H, 3.60; Cl, 23.01. Found: C, 44.57; H, 3.59; Cl, 23.22.

2,3,4,6-Tetra-O-chloroacetyl-1-O-(3,4,5-trimethoxybenzoyl)-D-glucopyranose (5b). (a) A solution of 2c (1.09 g, 2.23 mmol), 3b (0.62 g, 2.9 mmol) and boron trifluoride etherate (0.38 mL, 3.03 mmol) in dichloromethane (15 mL) was stirred at room temp for 30 h. Work up as described for 5a (c) gave a 90:10 α/β -mixture (¹H NMR) of 5b (0.79 g, 52%). Chromatography (solvent A, 5:1) gave α -5b (0.54 g, 36%) as a colourless glass, [α]_D +51.8° (c 1.0, chloroform). ¹H NMR δ 6.55 (d, 1 H, $J_{1,2} = 3.4$ Hz, H-1), 5.80 (t, 1 H, $J_{2,3} = J_{3,4} = 9.9$ Hz, H-3), 5.30 (dd, 1 H, H-2), 5.26 (t, 1 H, $J_{4,5} = 9.9$ Hz, H-4), 4.40 (dd, 1 H, $J_{5,6a} = 2.6$ Hz, $J_{6a,6b} = -12.8$ Hz, H-6a), 4.32 (dd, 1 H, $J_{5,6b} = 2.4$ Hz, H-6b), 4.24-4.19 (m, 1 H, H-5), 4.13, 4.04, 4.00, 4.06 (3 x s, d, 4 x 2 H, CH₂Cl), 3.96, 3.95 (2 x s, 6 H, 3 H, OCH₃); ¹³C NMR δ 89.3 (C-1), 70.9, 70.6, 69.4, 69.3 (C-2,3,4,5), 62.8 (C-6), 61.0, 56.5 (1 C, 2 C, OCH₃), 40.5, 40.2 (2 x 2 C, CH₂Cl). Anal. Calcd for C₂₄H₂₆Cl₄O₁₄: C, 42.37; H, 3.85; Cl, 20.85. Found: C, 42.58; H, 4.04; Cl, 20.79.

(b) A suspension of 2g (912.0 mg, 1.66 mmol) and 4b (811.9 mg, 2.54 mmol) in dichloromethane (8 mL) was stirred at room temp for 3 h. Work up as described for 5a (e) gave β -5b (723.4 mg, 64%) as a colourless foam, [α]_D -26.2° (*c* 1.0, chloroform). ¹H NMR δ 5.91 (d, 1 H, $J_{1,2} = 7.7$ Hz, H-1), 5.55-5.41 (m, 2 H, H-2,3), 5.29 (t, 1 H, $J_{3,4} = J_{4,5} = 9.4$ Hz, H-4), 4.45 (dd, 1 H, $J_{5,6a} = 4.2$ Hz, $J_{6a,6b} = -12.5$ Hz, H-6a), 4.34 (dd, 1 H, $J_{5,6b} = 2.1$ Hz, H-6b), 4.12, 4.03 (2 x s, 2 x 2 H, CH₂Cl), 4.05, 4.00 (2 x d, 2 x 2 H, CH₂Cl), 3.92, 3.91 (2 x s, 3 H, 6 H, OCH₃), 3.88-3.78 (m, 1 H, H-5); ¹³C NMR δ 92.0 (C-1), 73.4, 72.2, 71.3 (C-2,3,5), 69.2 (C-4), 62.7 (C-6), 61.0 (OCH₃), 56.3 (2 C, OCH₃), 40.5, 40.2 (1 C, 3 C, CH₂Cl).

Anal. Calcd for C₂₄H₂₆Cl₄O₁₄: C, 42.37; H, 3.85; Cl, 20.85. Found: C, 42.66; H, 4.03; Cl, 20.67.

2,3,4,6-Tetra-O-chloroacetyl-1-*O*-(2-methyl-4-pentenoyl)-D-glucopyranose (5c). (a) A solution of 2c (1.02 g, 2.08 mmol), 3c (0.3 g, 2.59 mmol) and boron trifluoride etherate (0.4 mL, 3.18 mmol) in dichloromethane (16 mL) was stirred at room temp for 24 h. Work up as described for 5a (c) gave a 72:28 α/β -mixture (¹H NMR) of 5c (0.73 g, 60%). Chromatography (solvent A, 5:1) gave α -5c (0.41 g, 34%) as a colourless foam, [α]_D +73.2° (*c* 1.3, chloroform). ¹H NMR δ 6.38 (d, 1 H, $J_{1,2}$ = 3.7 Hz, H-1), 5.86-5.69 (m, 1 H, H₂C=CH), 5.59 (t, 1 H, $J_{2,3} = J_{3,4} = 9.8$ Hz, H-3), 5.24 (t, 1 H, $J_{4,5} = 9.8$ Hz, H-4), 5.22-4.93 (m, 3 H, H-2, H_2 C=CH), 4.37 (dd, 1 H, $J_{5,6a} = 4.1$ Hz, $J_{6a,6b} = -12.5$ Hz, H-6a), 4.32-4.17 (m, 2 H, H-5,6b), 4.12, 4.02, 4.05, 3.98 (2 x s, 2 x d, 4 x 2 H, CH₂Cl); ¹³C NMR δ 134.6 (H₂C=CH), 117.6 (H₂C=CH), 88.4 (C-1), 71.0, 70.5, 69.5, 69.1 (C-2,3,4,5), 62.8 (C-6), 40.5, 40.3, 40.2, 40.1 (CH₂Cl), 39.1 (*C*H-CH₃), 37.4 (CH₂), 16.5 (CH-CH₃).

Anal. Calcd for C₂₀H₂₄Cl₄O₁₁: C, 41.26; H, 4.15; Cl, 24.36. Found: C, 41.03; H, 4.18; Cl, 23.94.

(b) A suspension of 2g (685.0 mg, 1.25 mmol) and 4c (465.3 mg, 2.11 mmol) in dichloromethane (20 mL) was stirred at room temp for 24 h. Work up as described for 5a (e) gave β -5c (508.4 mg, 70%) as a colourless foam, [α]_D +6.8° (*c* 1.0, chloroform). ¹H NMR δ 5.82 (d, 1 H, $J_{1,2} = 8.3$ Hz, H-1), 5.74-5.62 (m, 1 H, H₂C=CH), 5.40 (t, 1 H, $J_{2,3} = J_{3,4} = 9.4$ Hz, H-3), 5.28-5.19 (m, 2 H, H-2,4), 5.10-5.07, 5.04-5.03 (2 x m, 2 x 1 H, H_2 C=CH), 4.41 (dd, 1 H, $J_{5,6a} = 4.3$ Hz, $J_{6a,6b} = -12.5$ Hz, H-6a), 4.30 (dd, 1 H, $J_{5,6b} = 2.3$ Hz, H-6b), 4.12, 4.00, 3.99, 4.03 (3 x s, d, 4 x 2 H, CH₂Cl), 3.97-3.93 (m, 1 H, H-5); ¹³C NMR δ 134.5 (H₂C=CH), 117.5 (H₂C=CH), 91.1 (C-1), 73.8, 72.1, 71.3 (C-2,3,5), 69.1 (C-4), 62.7 (C-6), 40.5, 40.2 (1 C, 3 C, CH₂Cl), 39.0 (*C*H-CH₃), 37.2 (CH₂), 16.2 (CH-CH₃).

Anal. Calcd for C₂₀H₂₄Cl₄O₁₁: C, 41.26; H, 4.15; Cl, 24.36. Found: C, 40.92; H, 4.19; Cl, 23.62.

2,3,4,6-Tetra-O-chloroacetyl-1-O-oleoyl-β-D-glucopyranose (5d). A suspension of 2g (1.45 g, 2.64 mmol) and 4d (1.27 g, 3.26 mmol) in dichloromethane (50 mL) was stirred at room temp for 72 h. Work up as described for 5a (e) and chromatography (solvent A, 10:1) gave 5d (1.50 g, 75%) as a wax like solid, $[\alpha]_D$ -8.7° (*c* 1.0, chloroform). ¹H NMR δ 5.79 (d, 1 H, $J_{1,2} = 8.1$ Hz, H-1), 5.54 (d, 2 H, J = 7.9 Hz, *CH=CH*), 5.41-5.30 (m, 2 H, H-2,3), 5.22 (t, 1 H, $J_{3,4} = J_{4,5} = 9.4$ Hz, H-4), 4.51-4.39 (m, 1 H, H-6a), 4.30 (dd, 1 H, $J_{5,6b} = 2.0$ Hz, $J_{6a,6b} = -12.4$ Hz, H-6b), 4.15, 4.12, 4.03, 4.00 (4 x s, 4 x 2 H, CH₂Cl), 3.96-3.75 (m, 1 H, H-5); ¹³C NMR δ 130.1, 129.7 (*C*H=*C*H), 93.6 (C-1), 73.9, 72.5, 71.3, 69.8 (C-2,3,4,5), 62.7 (C-6), 40.9, 40.5, 40.2 (2 x 1 C, 2 C, CH₂Cl).

Anal. Calcd for C₃₂H₄₈Cl₄O₁₁: C, 51.21; H, 6.45; Cl, 18.89. Found: C, 51.00; H, 6.45; Cl, 18.39.

2,3,4,6-Tetra-O-chloroacetyl-1-O-(4-chloroacetoxycinnamoyl)-D-glucopyranose (5e). A solution of 2c (557.3 mg, 1.14 mmol), 3e (346.2 mg, 1.44 mmol) and boron trifluoride etherate (0.2 mL, 1.59 mmol) in dichloromethane (20 mL) was stirred at room temp for 56 h. Work up as described for 5a (c) gave a 77:23 α/β -mixture (¹H NMR) of 5e (429.5 mg, 51%). Chromatography (solvent A, 5:1) gave α -5e (219.7 mg, 27%) as a colourless oil, [α]_D +43.3° (*c* 0.7, chloroform). ¹H NMR δ 7.68 (d, 1 H, J = -16.5 Hz, CH=CHCOO), 6.28 (d, 1 H, CH=CHCOO), 6.27 (d, 1 H, $J_{1,2}$ = 4.1 Hz, H-1), 5.68 (t, 1 H, $J_{2,3} = J_{3,4} = 9.8$ Hz, H-3), 5.23 (t, 1 H, $J_{4,5} = 9.8$ Hz, H-4), 5.15 (dd, 1 H, H-2), 4.76, 4.14, 4.13, 4.09, 4.08 (5 x s, 5 x 2 H, CH₂Cl), 4.45-4.16 (m, 3 H, H-5,6a,6b); ¹³C NMR δ 146.3 (CH=CHCOO), 116.0 (CH=CHCOO), 89.1 (C-1), 72.1, 71.5, 70.0, 69.1 (C-2,3,4,5), 62.5 (C-6), 41.6, 40.6, 40.2 (1 C, 2 x 2 C, CH₂Cl).

Anal. Calcd for C₂₅H₂₃Cl₅O₁₃: C, 42.37; H, 3.27. Found: C, 42.69; H, 3.38.

2,3,4,6-Tetra-O-chloroacetyl-1-O-cinnamoyl-\beta-D-galactopyranose (7a). A suspension of 6 (1.03 g, 1.88 mmol) and 4a (0.71 g, 2.8 mmol) in dichloromethane (20 mL) was stirred at room temp for 40 h. Work up as described for 5a (e) gave 7a (0.81 g, 70%) as a colourless foam, [α]_D -3.7° (*c* 1.0, chloroform). ¹H NMR δ 7.78 (d, 1 H, *J* = -16.0 Hz, *CH*=CHCOO), 6.42 (d, 1 H, CH=CHCOO), 5.93 (d, 1 H, *J*_{1,2} = 8.2 Hz, H-1), 5.57-5.48 (m, 2 H, H-2,4), 5.34 (dd, 1 H, *J*_{2,3} = 10.4 Hz, *J*_{3,4} = 3.4 Hz, H-3), 4.37-4.27 (m, 3 H, H-5,6a,6b), 4.22, 4.08, 4.02 (3 x s, 2 x 2 H, 4 H, CH₂Cl); ¹³C NMR δ 148.1 (*C*H=CHCOO), 115.7 (CH=*C*HCOO), 91.7 (C-1), 72.0, 71.2, 69.1, 68.6 (C-2,3,4,5), 62.1 (C-6), 40.5, 40.3 (2 x 2 C, CH₂Cl).

Anal. Calcd for $C_{23}H_{22}Cl_4O_{11}$: C, 44.83; H, 3.60; Cl, 23.01. Found: C, 44.63; H, 3.60; Cl, 22.34.

2,3,4,6-Tetra-O-chloroacetyl-1-O-(3,4,5-trimethoxybenzoyl)- β -D-galactopyranose (7b). A suspension of 6 (911.6 mg, 1.66 mmol) and 4b (638.2 mg, 2.00 mmol) in dichloromethane (20 mL) was stirred at room temp for 2 h. Work up as described for 5a (e) gave 7b (627.3 mg, 56%) as a colourless foam, $[\alpha]_D$ -16.4° (c 1.0, chloroform). ¹H NMR δ 5.89 (d, 1 H, $J_{1,2} = 8.3$ Hz, H-1), 5.61-5.57 (m, 2 H, H-2,4), 5.37 (dd, 1 H, $J_{2,3} = 10.5$ Hz, $J_{3,4} = 3.4$ Hz, H-3), 4.39-4.26 (m, 3 H, H-5,6a,6b), 4.22, 4.08, 4.03, 4.00 (3 x s, d, 4 x 2 H, CH₂Cl), 3.93 (s, 9 H, OCH₃); ¹³C NMR δ 92.4 (C-1), 71.6, 71.3, 69.0, 68.4 (C-2,3,4,5), 61.9 (C-6), 61.0, 56.3 (1 C, 2 C, OCH₃), 40.4, 40.3 (2 x 2 C, CH₂Cl).

Anal. Calcd for $C_{24}H_{26}Cl_4O_{14}$: C, 42.37; H, 3.85; Cl, 20.85. Found: C, 42.61; H, 3.84; Cl, 20.30.

2,3,4,6-Tetra-O-chloroacetyl-1-*O*-(2-methyl-4-pentenoyl)- β -D-galactopyranose (7c). A suspension of 6 (702.0 mg, 1.28 mmol) and 4c (473.6 mg, 2.14 mmol) in dichloromethane (20 mL) was stirred at room temp for 24 h. Work up as described for 5a (e) gave 7c (410.8 mg, 55%) as a colourless foam, $[\alpha]_D$ +14.1° (*c* 1.0, chloroform). ¹H NMR δ 5.79 (d, 1 H, $J_{1,2} = 8.1$ Hz, H-1), 5.75-5.60 (m, 1 H, H₂C=CH), 5.53-5.48 (m, 1 H, H-4), 5.41 (dd, 1 H, $J_{2,3} = 10.3$ Hz, H-2), 5.28 (dd, 1 H, $J_{3,4} = 3.3$ Hz, H-3), 5.11-5.07, 5.04-5.03 (2 x m, 2 H, H_2 C=CH), 4.38-4.25 (m, 3 H, H-5,6a,6b), 4.20, 4.07, 4.00, 4.01 (3 x s, d, 4 x 2 H, CH₂Cl); ¹³C NMR δ 134.6 (H₂C=CH), 117.4 (H₂C=CH), 91.5 (C-1), 72.0, 71.1, 69.0, 68.5 (C-2,3,4,5), 62.0 (C-6), 40.4, 40.2 (2 x 2 C, CH₂Cl), 39.0 (CH-CH₃), 37.2 (CH₂), 16.2 (CH-CH₃).

Anal. Calcd for C₂₀H₂₄Cl₄O₁₁: C, 41.26; H, 4.15. Found: C, 40.99; H, 4.17.

2,3,4,6-Tetra-O-chloroacetyl-1-O-oleoyl-\beta-D-galactopyranose (7d). A suspension of 6 (1.45 g, 2.63 mmol) and 4d (1.28 g, 3.3 mmol) in dichloromethane (50 mL) was stirred at room temp for 72 h. Work up as described for 5a (e) gave 7d (1.36 g, 69%) as a highly viscous oil, $[\alpha]_D$ +9.6° (c 1.3, chloroform). ¹H NMR δ 5.79 (d, 1 H, $J_{1,2} = 8.1$ Hz, H-1), 5.52-5.49 (m, 1 H, H-4), 5.40 (dd, 1 H, $J_{2,3} = 10.3$ Hz, H-2), 5.36-5.30 (m, 2 H, CH=CH), 5.26 (dd, 1 H, $J_{3,4} = 3.3$ Hz, H-3), 4.37-4.22 (m, 3 H, H-5,6a,6b), 4.19, 4.06, 4.00, 3.99 (4 x s, 4 x 2 H, CH₂Cl); ¹³C NMR δ 130.1, 129.7 (CH=CH), 91.4 (C-1), 72.0, 71.2, 69.1, 68.5 (C-2,3,4,5), 62.0 (C-6), 40.4, 40.2 (2 x 2 C, CH₂Cl).

Anal. Calcd for C₃₂H₄₈Cl₄O₁₁: C, 51.21; H, 6.45. Found: C, 51.27; H, 6.50.

1-O-Acyl-D-glycopyranoses (8, 9). General Procedure. A solution of 5 and 7, respectively and 1-piperidinethiocarbamide¹³ (2 equivalents per chloroacetyl group to be removed) in methanol/dichloromethane (1:1) for compd 5a or methanol/chloroform (1:1) for all other compds was stirred at room temp for 18-24 h until all starting material had reacted (TLC, solvent *B*, 10:1). Concentration and chromatography (solvent *B*, 10:1) gave 8 and 9, respectively.

1-O-Cinnamoyl-β-D-glucopyranose (8a). According to the General Procedure, 5a (0.66 g, 1.38 mmol) afforded 8a (0.25 g, 58%) as an amorphous solid, $[\alpha]_D$ -8.2° (c 0.9, methanol).¹³C NMR δ 147.6 (CH=CHCOO), 118.2 (CH=CHCOO), 95.9 (C-1), 78.8, 77.9, 74.0, 71.0 (C-2,3,4,5), 62.3 (C-6).

Anal. Calcd for C₁₅H₁₈O₇: C, 58.06; H, 5.85. Found: C, 57.71; H, 6.12.

1-*O*-(3,4,5-Trimethoxybenzoyl)-β-D-glucopyranose (8b). According to the General Procedure, β-5b (313.5 mg, 0.46 mmol) afforded 8b (104.8 mg, 61%) as a colourless glass, $[\alpha]_D$ -7.1° (*c* 0.9, methanol).¹³C NMR δ 97.8 (C-1), 80.2, 79.3, 75.4, 72.4 (C-2,3,4,5), 63.7 (C-6), 62.6, 58.2 (1 C, 2 C, OCH₃).

DCI-MS (NH₃): m/z = 375 (MH⁺), 392 (MH⁺+NH₃).

1-O-(2-Methyl-4-pentenoyl)-β-D-glucopyranose (8c). According to the General Procedure, β-5c (146.8 mg, 0.25 mmol) afforded 8c (37.6 mg, 56%) as a colourless hygroscopic solid, $[\alpha]_D$ +9.2° (c 0.7, methanol). ¹³C NMR δ 136.6 (H₂C=CH), 117.5 (H₂C=CH), 95.8 (C-1), 78.9, 78.1, 74.0, 71.1 (C-2,3,4,5), 62.4 (C-6), 40.3 (CH-CH₃), 38.5 (CH₂), 16.5 (CH-CH₃).

DCI-MS (NH₃): $m/z = 276 (M^+)$, 277 (MH⁺), 294 (MH⁺+NH₃).

1-O-Oleoyl-β-D-glucopyranose (8d). According to the General Procedure, β-5d (609.2 mg, 0.81 mmol) afforded 8d (185.6 mg, 52%) as a colourless hygroscopic solid, $[\alpha]_D$ +15.4° (*c* 0.6, methanol). ¹³C NMR δ 130.7, 130.6 (*C*H=*C*H), 98.1 (C-1), 77.9, 77.8, 76.2, 71.7 (C-2,3,4,5), 62.6 (C-6).

DCI-MS (NH₃): m/z = 444 (M⁺).

1-*O*-Cinnamoyl-β-D-galactopyranose (9a). According to the General Procedure, 7a (367.4 mg, 0.6 mmol) afforded 9a (118.4 mg, 64%) as a colourless foam, $[\alpha]_D$ +17.8° (*c* 0.6, methanol). ¹³C NMR δ 147.6 (*C*H=CHCOO), 119.7 (CH=*C*HCOO), 97.9 (C-1), 78.9, 76.2, 72.7, 71.5 (C-2,3,4,5), 63.6 (C-6).

Anal. Calcd for C₁₅H₁₈O₇: C, 58.06; H, 5.85. Found: C, 57.70; H, 5.98.

1-*O*-(3,4,5-Trimethoxybenzoyl)-β-D-galactopyranose (9b). According to the General Procedure, 7b (146.3 mg, 0.22 mmol) afforded 9b (45.3 mg, 55%) as a colourless hygroscopic solid, $[\alpha]_D$ +12.4° (*c* 0.6, methanol). ¹³C NMR δ 96.8 (C-1), 78.8, 77.5, 73.7, 72.7 (C-2,3,4,5), 64.7 (C-6), 59.3, 54.8 (1 C, 2 C, OCH₃).

DCI-MS (CH₄): m/z = 374 (M⁺), 375 (MH⁺).

ACKNOWLEDGMENT

We thank Prof. Dr. Dr. h.c. F. Effenberger for helpful discussions and for providing the laboratory facilities. We also thank Dr. W. Rozdzinsky for performing the elemental analyses and Dr. P. Fischer and J. Rebell for recording the NMR spectra. We also thank the Hoechst AG, Frankfurt for a gift of α -D-glucopyranosyl fluoride. This work was financially supported by the Deutsche Forschungsgemeinschaft.

REFERENCES

- 1. T. Ziegler, Liebigs Ann. Chem., 1990, 1125.
- 2. C. A. A. van Boeckel, T. Beetz, Tetrahedron Lett., 24, 3755 (1983).
- 3. Y. Nishikawa, K. Yoshimoto, K. Ashizawa, T. Ikekawa, *Chem. Pharm. Bull.*, 29, 880 (1981); and references cited therein.
- 4. H. Pfänder, Pure & Appl. Chem., 51, 565 (1979).
- 5. D. Plusquellec, F. Roulleau, F. Bertho, M. Lefeuvre, E. Brown, *Tetrahedron*, 42, 2457 (1986).
- 6. K. Yoshimoto, M. Taru, Y. Tsuda, Tetrahedron Lett., 24, 2779 (1983).
- 7. H. A. Vaccaro, R. A. Rivero, A. B. Smith, Tetrahedron Lett., 30, 1465 (1989).
- 8. A. Lubineau, E. Meyer, P. Place, Carbohydr. Res., 228, 191 (1992).
- 9. G. J. F. Chittenden, H. Regeling, Recl. Trav. Chim. Pays-Bas, 106, 44 (1987).
- 10. M. Bertolini, C. P. J. Glaudemans, Carbohydr. Res., 15, 263 (1970).
- 11. T. Ziegler, P. Kováč, C. P. J. Glaudemans, Carbohydr. Res., 194, 185 (1989).
- 12. P. Konradsson, U. E. Udodong, B. Fraser-Reid, *Tetrahedron Lett.*, **31**, 4313 (1990).
- 13. H. Hartmann, I. Reuther, J. Prakt. Chem., 315, 144 (1973).
- 14. W. Steglich, H. G. Batz, Angew. Chem., Int. Ed. Engl., 10, 75 (1971).
- 15. E. Fischer, K. Delbrück, Ber. Dtsch. Chem. Ges., 42, 1476 (1909).
- 16. R. J. Ferrier, R. H. Furneaux in Meth. Carbohydr. Chem., Academic Press, New York 1980, vol. VIII, p 251.
- 17. R. U. Lemieux, Can. J. Chem, 29, 1079 (1951).